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Research article

Testosterone variation in a semi-captive population of Asian elephants in Myanmar

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Hormones are known to be involved in life-history trade-offs as systemic signals that establish functional links among traits and regulate key behavioural and physiological transitions between states in organisms. Although major functions of many steroid hormones such as testosterone are conserved among vertebrates, circulating concentrations vary widely both within and across species, and the degree to which observed hormone concentrations mediate life-history responses to environmental variation is less understood. In this study, we investigated how faecal testosterone metabolite (FTM) concentrations varied with extrinsic and intrinsic factors. To do so, we took advantage of a 6-year period of longitudinal sampling of FTM, indicators of stress and oxidative status in a semi-captive population of Asian elephants (n = 3163 samples from 173 individuals) in Myanmar. We determined how the variation in FTM is associated with age, sex, origin (captive-born or wild-caught), seasonality of the environment, individual stress level [measured by faecal glucocorticoid metabolite (FGM) and heterophil to lymphocyte ratio (H/L)] and oxidative status (reactive oxygen metabolite concentrations and superoxide dismutase activity). We reported that FTM increased with age from juvenile to adulthood for both sexes, with higher FTM concentrations in males than females. Moreover, elephants showed significantly higher FTM concentrations during the hot season and monsoon than in the cold season. However, for the physiological indicators, we found contrasting results. While FTM concentrations were strongly positively correlated with FGM concentrations, FTM concentrations were not related to H/L ratios. Finally, we found no relationship between FTM and the oxidative status of individuals. Our study provides new insights on the factors associated with variation in testosterone concentrations—a key hormone for reproduction and fitness of individuals—in Asian elephants living in their natural environment, which has relevance for effective conservation measures of this endangered species.

Lay Summary

In this study, we investigated how the concentration of testosterone, a key hormone for reproduction and fitness of individuals, varied with extrinsic and intrinsic factors in a semi-captive population of Asian elephants. We determined its association with age, sex, origin of elephants, seasonal variation, individuals' stress level and individuals' oxidative status.

Key words: Asian elephant, environmental factors, glucocorticoid, H/L ratio, hormones, oxidative stress, testosterone

Abbreviations: T: testosterone; FTM: faecal testosterone metabolite; FGM: faecal glucocorticoid metabolite; H/L ratio: heterophils and lymphocytes ratio; ROMs: reactive oxygen metabolites; SOD: superoxide dismutase.

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Introduction

Testosterone, a steroid hormone produced by gonads, is central in the development of reproductive organs and spermatogenesis, and secondary sexual characteristics involved in mating and reproduction (Ketterson et al., 1996; Ketterson and Val, 1999; Wingfield et al., 2001). Testosterone supports phenotypic traits that enhance attractiveness and increase mating effort (Wingfield, 1984; Ketterson and Val, 1999: Stoehr and Hill, 2000: McGlothlin et al., 2007: Mills et al., 2009; Negro et al., 2010). However, the secretion of testosterone also induces costs for the organisms. High testosterone concentrations can decrease survival rate (Marler and Moore, 1988; Dufty, 1989; Reed et al., 2006; Mills et al., 2009) and parental care (Silverin, 1980; Hegner and Wingfield, 1987; Oring et al., 1989; Stoehr and Hill, 2000; O'Neal et al., 2008; Veiga et al., 2008) and can negatively impact female reproductive success and fitness as observed in birds (Rutkowska et al., 2005; O'Neal et al., 2008; Veiga et al., 2008). In mammals, an excess of testosterone during prenatal development can cause physiological disruption in adulthood, including disruptions of reproductive processes (Padmanabhan et al., 2006). Therefore, testosterone has a direct influence on the fitness of individuals (Wingfield et al., 2001).

While the role of steroid hormones is conserved among vertebrates, testosterone concentrations vary both between and within individuals (Kempenaers et al., 2008). Males display higher concentrations of testosterone than females in mammals (Rasmussen et al., 1984; Glickman et al., 1987, 1992; Holekamp and Talamantes, 1991; Whitworth et al., 1999; Goymann et al., 2001; Dittami et al., 2008; Burgess et al., 2012), and the concentrations vary according to life stage or lifespan (Kempenaers et al., 2008). In some long-lived mammals, the mean testosterone concentration increases with age, as in male elephants (Brown et al., 2007), but decreases in very old individuals (Thongtip et al., 2008). Similarly, in primates, androgen hormone levels increase with age (Martin et al., 1977), with the highest testosterone concentrations observed at maturity, after which they decline (Beehner et al., 2005, 2009; Seraphin et al., 2008) or reach a plateau (Crawford et al., 1997). However, age-related variation in testosterone concentrations among taxa still needs to be assessed, especially in females, since studies to date have mainly focused on males.

Beyond individual characteristics, testosterone secretion is influenced by extrinsic factors from the environment,

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sometimes reflecting the adaptive mechanisms of an individual in a changing environment (Wingfield and Kenagy, 1991; Astheimer et al., 1995). Seasonality, marked by variations in climate, food availability, disease prevalence and predation, is an important ecological factor known to affect testosterone concentrations (Whitehead and McEwan, 1973; Pelletier et al., 1982; Orlando et al., 2003; Gündoğan, 2007; Sun et al., 2011). In captive Asian elephants in Thailand, for example, males display seasonal variation in T, with higher testosterone concentrations during the cold season and monsoon, when food availability is higher than in the hot season (Thongtip et al., 2008; Norkaew et al., 2019). Hence, environmental variation can affect hormonal concentrations, and it is essential to consider this in the study of testosterone concentration variation, as intrinsic factors can be confounded with environmental effects (Kempenaers et al., 2008).

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Some environmental fluctuations (e.g. climate, food availability) cause an activation of the hypothalamic-pituitaryadrenal (HPA) axis with the production of glucocorticoids, resulting in the suppression of the hypothalamic-pituitarygonadal (HPG) axis, and thus lower testosterone levels. This decrease in testosterone secretion following a stress event has been evidenced in male vertebrates, such as humans (Matsumoto et al., 1970; Kreuz et al., 1972; Aakvaag et al., 1978), tree lizards (Moore et al., 1991), rats (Gray et al., 1978) and silver foxes (Moe and Bakken, 1996), and could disrupt or suppress reproduction (Sapolsky, 2002; Wingfield and Sapolsky, 2003). Concentrations of glucocorticoids, including corticosterone, are often used as an indicator of stress in various mammalian species (Keay et al., 2006). Similarly, the ratio between two types of white blood cells-heterophils (or neutrophils in most mammals) and lymphocytes-referred to as heterophil to lymphocyte ratio (H/L) ratio, is considered a fairly reliable stress marker, with decreasing lymphocytes in blood and increasing neutrophils or heterophils in response to stress (Gross and Siegel, 1983; McFarlane and Curtis, 1989; Vleck et al., 2000). These two physiological responses to a stressor are correlated (Seltmann et al., 2020), but they do not have the same role and differ temporally in their response to stress (Davis and Maney, 2018). They might offer ways of elucidating the interplay between endocrinological and haematological responses, and how they might interact in phenotypic plasticity.

Another physiological stress consequence is oxidative stress, manifested by overproduction and accumulation of free radicals as a result of an imbalance between the

production of reactive oxygen species (ROS) and the activation of various antioxidant defences (Halliwell, 1994). Previous studies, for example, on rats (Chainy et al., 1997; Pansarasa et al., 2002), rabbits (Aydilek et al., 2004), humans (Zhu et al., 1997) and birds (Alonso-Alvarez et al., 2007), showed that oxidative stress increased concomitantly with increasing concentrations of testosterone. Testosterone can increase ROS production and impair the antioxidant system through genomic and non-genomic effects (see for example Chainy et al., 1997; Cruz-Topete et al., 2020). However, other studies have obtained different or even contradictory results. In a study on White's skinks, Isaksson et al. (2011) found that an aggressive behavioural phenotype, rather than testosterone concentrations, was associated with higher oxidative stress. Furthermore, some medical studies found a protective effect of testosterone reducing oxidative stress in the brain (Túnez et al., 2007; Meydan et al., 2010; Toro-Urrego et al., 2016; Yan et al., 2017; Cai and Li, 2020). Hence, further studies on a wider range of species investigating the link between testosterone variation and the oxidative status of individuals are needed to better understand the influence of testosterone on the fitness of individuals.

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Although there is extensive research on the role of testosterone, we know far less about the determinants of its variation and the mechanisms mediating the hormonal response to environmental changes. In this study, we investigated the intrinsic and extrinsic determinants of faecal testosterone metabolite (FTM) variation in the Asian elephant measured from faecal samples, a non-invasive method used in numerous studies including on Asian elephants (Seltmann et al., 2020; de Andrés et al., 2021; Glaeser et al., 2021; LaDue et al., 2022, 2023; Ukonaho et al., 2023). Previous studies on the testosterone endocrinology of the Asian elephant focused primarily on males during the rutting period called musth, characterized by a peak of testosterone concentrations, high aggressiveness and high sexual activity (Rasmussen and Perrin, 1999; Brown et al., 2007; Yon et al., 2007a, 2007b, 2008; Kumar et al., 2014). In contrast, very few studies have examined testosterone variation at a basal concentration in both sexes. We took advantage of our unique dataset on a semi-captive population of elephants in Myanmar, working during the day in timber camps managed by the Myanma Timber Enterprise (MTE), but free roaming at night in the surrounding forest. We collected 3163 FTM samples from 173 elephants over 6 years and analysed the association between FTM variation and individual characteristics (age, sex, wild-caught or captive-born origin), the environment (seasonal effect) and potential indicators of physiological stress [faecal glucocorticoid metabolite (FGM) concentrations and H/L ratio] and the possible consequences of FTM variation on the oxidative status of individuals [reactive oxygen metabolite (ROM) concentrations and superoxide dismutase (SOD) activity]. Determining the link between ecological and physiological factors with testosterone level, a crucial hormone for reproduction and fitness, is important to understanding the survival and reproduction patterns of the Asian

elephant. By identifying the factors affecting testosterone variation in Asian elephants, our study will provide essential information to develop targeted conservation strategies and may offer management solutions for this endangered species. We expected the following: (1) higher FTM in males and (2) sexually mature individuals, but lower in the oldest elephants; (3) a seasonal effect, known to affect health, markers of stress and mortality risks in our population (Mumby et al., 2013a, 2013b; dos Santos et al., 2020; Ukonaho et al., 2023), with higher FTM concentrations during monsoon due to higher food availability, which we expect would lead to a greater activity of the HPG axis. For the underlying physiological factors, we hypothesized (4) a suppressive effect of stress on FTM, thus a negative relationship between FGM concentrations and H/L ratios with FTM concentrations. We also predicted an origin-related variation in FTM concentrations, with lower concentrations for wild-caught than captive-born elephants as a result of the higher stress levels they experience in captivity (Lahdenperä et al., 2018; Jackson et al., 2019). Finally, we expected (5) a negative effect of higher FTM concentrations on the oxidative status of individuals, i.e. elevated biomarkers of oxidative stress.

Materials and Methods

Study model

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We studied 173 semi-captive elephants from Myanmar, owned by the governmental MTE and employed in the timber industry (Schmidt and Mar, 1996). These elephants were managed during the day by their allocated handler (called a mahout) in logging camps to handle and pull logs, but they were free to roam at night and outside of working hours (5 to 8 hours during the day) in the surrounding forests. Most of these timber elephants were captive-born, but some had been captured from the wild. Since 1968, the capture of wild elephants was restricted to elephants of at least \sim 5 years of age in Myanmar, and it was banned more than two decades ago (Schmidt and Mar, 1996; Lahdenperä et al., 2018). Asian elephants can reproduce all year round, as female ovulation and musth in males can occur in any season (Fowler and Mikota, 2006; Brown et al., 2007; Hildebrandt et al., 2011). Reproduction in logging elephants is not managed by humans, and they can breed with wild elephants when released into the forest. Hence, these MTE elephants exhibit many foraging and breeding patterns comparable to those of wild elephants and are termed 'semi-captive' (Robinson et al., 2012; Lahdenperä et al., 2018).

Each of these semi-captive elephants was marked with a unique identification (ID) number and was regularly checked from birth to death by an MTE veterinarian, who recorded a wide range of life-history information in the elephant's logbook, such as their origin (captive-born or wild-caught), health status, morphological data, pedigree (mother ID) and dates of birth and death. The exact dates of birth of wildcaught elephants were estimated, as they were unknown, in

contrast to captive-born elephants, for whom the specific day, month and year of birth was known. The approximate age of wild-caught elephants was determined by comparing their body height and mass to those of captive-born elephants of known age (Lahdenperä *et al.*, 2018; Lalande *et al.*, 2022). In any given year, elephants in Myanmar experience three distinct seasons: cold season (October–January), hot season (February–May) and monsoon (June–September). Traditionally, elephants work during monsoon and cold seasons and rest during the hot season (Mumby *et al.*, 2013a, 2015; Lahdenperä *et al.*, 2018), but none of the elephants included in the current study were involved in heavy work at time of the sampling.

Determination of age classes

The population of MTE elephants has a predicted median lifespan of 30.81 years for males and 44.73 years for females (Lahdenperä et al., 2018). The age of the elephants in the current study ranged from a few months to 69 years old for the females, and from a few months to 62 years old for the males. We created five different age classes for the elephants, which followed both the life history and the work life of the general population. These classes were calves-at-heel (0-4 years old), weaned calves (5-10 years old), adolescents (11-20 years old), prime-aged elephants (21-54 years old) and elderly elephants (55+ years old). MTE elephant calves stay with their mother from birth to 4-5 years old when they are weaned. The interbirth interval is around 4-5 years in Asian elephants (Sukumar, 2003; Lahdenperä et al., 2014). At 5 years old, calves are separated from their mother to be tamed (Crawley et al., 2020). The taming process lasts around 4 weeks, at the end of which each calf is assigned to an individual handler and continues to be trained (5-10 years old) without working duties yet. Between 11 and 20 years old, adolescent elephants conduct light work only. They usually reach sexual maturity at around 14-15 years. Prime-aged elephants aged 21-54 years old are all adults. The highest fertility of female elephants is between 20 and 50 years old and declines after 50 years old, with a peak in births occurring when females are aged 20-25 years old (Lahdenperä et al., 2014). These prime-aged elephants belong to the workforce, when they drag heavier logs during full-time work, before they retire from work at 55 years old. The workloads are imposed and regulated for all elephants by MTE (Mar et al., 2012).

Faecal sample collection, testosterone metabolite extraction and hormone assays

Collection and extraction

Our study was conducted in Kawlin logging camps, in Sagaing region of Myanmar. It includes 173 elephants longitudinally sampled for faecal samples once a month (when possible) across 6 years, from 2013 to 2018, corresponding to an average of 56.3 ± 10 faecal samples collected monthly and used in this study. These samples were collected just after

defecation in the morning, generally between 6 a.m. and 9 a.m., by local veterinarians as part of their regular health monitoring. For male elephants, the sample collection was conducted for safety reasons outside musth, a period of high sexual activity characterized by a peak of testosterone and glucocorticoid hormones, and high aggressiveness. An aliquot of exactly 4.5 g was taken from each faecal bolus. The samples were transferred to a cooler box for a maximum of 8 hours before being stored in a freezer at -20° C.

Previously validated faecal hormone assays (Watson et al., 2013; Kumar et al., 2014) were used to measure FTM and FGM concentrations. Faecal samples were analysed in 12 separate batches at the Veterinary Diagnostic Laboratory, Chiang Mai University, Thailand. The faecal samples were dried in a hot air oven at 50°C before extracting the testosterone metabolites using a boiling extraction method (Crawley *et al.*, 2021; Supanta et al., 2024). Samples (~ 0.1 g) were placed in glass tubes with 5 ml of 90% ethanol and extracted by boiling in a water bath for 20 minutes, with 100% ethanol added to maintain the volume at 5 ml. Samples were then centrifuged. the faecal supernatants were combined and dried in a 50°C water bath, reconstituted by vortexing for 1 minute in 3 ml of ethanol, dried down again and finally re-suspended in 1 ml of methanol. Samples were diluted 1:3 in assay buffer and stored at -20° C until analysis. We then quantified the concentrations of testosterone metabolites by enzyme immunoassay (EIA) validated for elephants (Kumar et al., 2014, see below for details), using a rabbit anti-testosterone polyclonal antibody (R156/7) with a concentration of 1:110 000 and a horseradish peroxidase-conjugated testosterone tracer (HRP) with a concentration of 1:20 000. The sample dilution of the faecal samples was 1:4.

The FTM data used in this study consisted of 3163 observations from 173 individuals including 106 females and 67 males (1 to 54 observations per individual with an average of 18). FTM concentrations ranged from 6.04 to 281.4 ng/g of dry faeces (with outliers removed, see Statistical Analyses section), and the age of sampled individuals represented a wide range from a few months old to 69 years old.

Plate coating with secondary antibody

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96-well plates were pre-coated with 150 μ l (10 μ g/ml) of coating antibody (goat anti-rabbit IgG) diluted in coating buffer (16 mM Na₂HPO₄, 1.6 mM NaH₂PO₄; pH 8.0), covered with a plate cover and then stacked and left for 24 hours at ambient temperature. The coating buffer was then removed from each plate, and 0.25 ml of blocking buffer (0.1% Tween 20, 8.12 mM sodium phosphate dibasic, 1.88 mM sodium phosphate monobasic, 15 mM sodium chloride, 0.009% Kathon CG/ICP, 29.21 mM sucrose) was dispensed into each well. Blocked plates were stacked and left for 4–24 hours at room temperature. The wells were then emptied, blotted dry, and the plates were transferred to a desiccator and left at ambient temperature until the inside humidity reached less than 20%. The dried plates were

then packaged individually and heat sealed inside labelled foil Ziploc bags before being stored at 4°C until used for EIAs.

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Enzyme immunoassay

The pre-coated goat anti-rabbit plates were used. Seventyfive microliters of assay buffer (0.1% Tween 20, 13.68 mM Trizma base, 86.3 mM Trizma hydrochloride, 0.15 M sodium chloride, 10 mM ethylenediaminetetraacetic acid (EDTA), 0.09% Kathon CG/ICP, 0.1% bovine serum albumin (BSA), Standard Grade pH 7.0) was added to the non-specific binding (NSB) wells, and 50 µl of assay buffer were added into the other wells to act as maximum binding wells (Bo or 0 pg/ml). Then, 50 µl of testosterone standards (17-hydroxy-4-androsten-3-one, Steraloids Cat. #A6950; range from 0.047 to 12 ng/ml), quality controls, and samples were each added into a well in the plate. Twenty-five microliters of HRP with a concentration of 1:20 000 was immediately added to each well, followed by 25 µl of testosterone polyclonal antibody (R156/7) with a concentration of 1:110000 added to all wells except the NSB wells. The plate was then covered with a plate sealer and shaken at ambient temperature for 2 hours. Each well was washed four times with wash buffer (1:20 dilution), and 100 µl of 3,3',5,5'-Tetramethylbenzidine (TMB) substrate solution (KPL TMB Microwell Peroxidase Substrate System 2-contents) was added to each well before incubation at ambient temperature for around 30 minutes without shaking. Finally, 50 µl of stop solution (2 M H₂SO₄) was added to each well, and the optical density generated from each well was read at 450 nm by a microplate reader. The assay had a sensitivity of 0.0567 ng/ml, with intraassay coefficients of variation (CVs) lower than 10% and inter-assay CV of 12.73%. The assay sensitivity refers to the lowest concentration that can be accurately analysed, which corresponds to the lowest standard concentration, calculated in Magellan Software using a 4PL curve.

FGM extraction and hormone assay

FGM concentrations were measured by EIA using an assay validated for use in Asian elephants (see above for plate coating and EIA protocol). The plate was filled with samples and corticosterone standards (Sigma, 27840; 50 µl; range from 0.078 to 20 ng/ml), followed by corticosterone-HRP (25 µl; 1:30 000) and polyclonal rabbit anti-corticosterone antibody (CJM006, Coralie Munro, UC Davis, CA) (25 µl; 1:100 000), and each plate was shaken at ambient temperature for 1 hour. Following Watson et al. (2013) protocol, we utilized Nunc MaxiSorp[®] plates, with substrate reagents at ambient temperature, and dark incubation conditions to optimize the results. The intra- and inter-assay CVs were lower than 10% and equal to 11.8%, respectively. The assay sensitivity was 0.099 ng/ml, and the absorbance was measured at 450 nm. Data consisted of 2964 observations from 171 individuals (1 to 51 observations per individual with an average of 17) ranging in age from 0 to 69 years old. Glucocorticoid metabolite concentrations ranged from 3.2 to 409.7 ng/g of dry faeces (similar to de Andrés *et al.*, 2021).

Blood samples collection

Blood samples were collected once per season (three times/year), corresponding to an average of 39.9 ± 19.6 blood samples collected per season and used in this study. Blood was collected from an ear vein of each elephant using a Vacuette[®] system (Greiner Bio-One, Kremsmünster, Austria) in the morning of non-workdays by local veterinarians as part of their routine health monitoring of each elephant for which they have been trained. Blood samples were collected into 8-ml tubes with the anticoagulant EDTA for measuring the H/L ratio, and in 9-ml serum separator tubes for oxidative stress measurement (dos Santos et al., 2020). The H/L ratio was measured the same day, within 0 to 8 hours after collection (see Methods below). The serum separator tubes were centrifuged (RCF-1320 g) for 20 minutes 3 to 6 hours after collection. Around 2 ml of serum was then pipetted into separate tubes and frozen at -20° C. The serum samples were later transported to the University of Turku in Finland and stored at -80° C until used for oxidative stress measurement.

Blood samples were collected less frequently than faecal samples, resulting in larger numbers of samples for the faecal hormone analyses and smaller sample numbers for the measurements of H/L ratio and oxidative stress. All sample sizes are summarized in Table A1 in the supplementary materials.

Evaluation of H/L ratio

From blood samples collected three times in 2016, 2017 and 2018 during the three seasons (hot, cold and monsoon) in Myanmar, we assessed the ratio between heterophils and lymphocytes (H/L), another indicator of stress and arousal, which correlates with glucocorticoid concentrations in elephants (Seltmann et al., 2020). Seltmann et al. (2020) recommend including behavioural indicators of stress, which we do not have in this study, to help interpret physiological markers of stress. To measure the H/L ratio, we used a validated protocol for Asian elephants (Seltmann et al., 2020). We kept the blood samples in 8-ml tubes containing the anticoagulant EDTA (dos Santos et al., 2020). We stained blood samples collected in EDTA with Romanowsky stain and manually counted the number of leucocytes from the blood smears. Then, we calculated the ratio of heterophils to lymphocytes counted on each blood smear. H/L ratios from the individuals used in this study ranged from 0.21 to 5.6. These data consist of 274 observations from 89 individuals (1 to 6 observations per individual with an average of 3), ranging in age from 4 to 69 years old.

Oxidative stress assessment

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To investigate the physiological consequences of FTM variation, we also measured markers of oxidative status of the

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elephants from blood samples collected from 2014 to 2018 in Myanmar. Two markers of oxidative status were evaluated in plasma: ROMs, indicators of oxidative damage, and the activity of SOD, indicative of antioxidant defence. These analyses were performed at the University of Turku, Finland.

Reactive oxygen metabolites

We measured the concentration of ROMs using a d-ROMs Test kit (Diacron International) according to methods reported by Costantini and Dell'Omo (2006). The d-ROMs test measures hydroperoxides (ROOH), the main compounds contributing to the oxidant ability of the plasma (Costantini, 2016). Each well of a 96-well microplate (SpectraPlateTM; PerkinElmer[®]) was filled with 5 µl of serum samples and 200 µl of "master mix" from the d-ROMs Test kit. The plates were shaken for 10 seconds and incubated at 37°C for 75 minutes before being read with a spectrophotometer (PerkinElmer[®]) at 510 nm. ROMs concentration is expressed as mg H₂O₂ equivalent/dl. The concentration of ROM measured in individuals in this study ranged from 4 to 30.5 H₂O₂/dl, and the intra-individual variation based on duplicates was $5.12 \pm 1.02\%$. These data consist of 356 observations from 81 individuals (1 to 11 observations per individual with an average of 4), ranging in age from 4 to 64 years old.

Superoxide dismutase

SOD is an antioxidant enzyme involved in the first step of the cascade catalysing the dismutation of superoxide anion free radical (O_2^-) into oxygen and hydrogen peroxide; this reduces the amount of O2⁻, which can damage cells at high concentrations. We measured SOD enzymatic activity from 25 µl of serum samples using a SOD activity kit (Enzo® Life Sciences, USA) following the manufacturer's instructions. The plates were read at 450 nm with a spectrophotometer (PerkinElmer®). This test quantifies in vitro the kinetics of inhibition in superoxide formation resulting from SOD antioxidant activity. SOD activity from the individuals used in this study ranged from 15.4 to 683.9 U/ml (units of enzymatic activity per milliliter), and the intra-individual variation based on duplicates was $23.3 \pm 4.64\%$. These data consist of 438 observations from 91 individuals (1 to 13 observations per individual with an average of 5), ranging in age from 4 to 65 years old.

Statistical analyses

We investigated the association of season, individual characteristics (age, sex, origin) and indicators of stress (FGM, H:L ratio, oxidative status) with FTM concentrations. All analyses were carried out on R software version 3.6.3 (R Core Team, 2022), and the graphical representations were constructed with 'ggplot2' package (Wickham, 2016). These figures show on the Y axis the values of FTM concentrations, in function of an explanatory variable, corrected for all covariates in the model. Correcting for covariates is a method well supported by other studies (e.g. Briga *et al.*, 2019; Briga and Verhulst,

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2021) to provide a figure that matches the tested model, and because covariates are accounted for, it prevents showing false positives or false negatives.

First, we analysed the distribution of FTM concentrations measured and the presence of potential outlier data points using *horn.outlier* function (package 'referenceIntervals'; Finnegan, 2015), which identifies data points outside 1.5*IQR (intequartile range) from the first or third quartile as an outlier.

We detected 48 outliers from 3211 observations (1.49% of all data points), which were removed from our analysis. The final dataset contained 3163 observations from 173 elephants. We applied a logarithmic transformation to the FTM values, to fit them to a normal distribution, before using linear mixed models (package 'lme4', Bates *et al.*, 2015), adjusted with the maximum likelihood method. We used the summary of the model to obtain the model parameters and *P* value of each variable. To compare the significance between the modalities of the factors in these models, we performed *post hoc* tests with the function 'diffIsmeans' from the package 'lmeTest' (Kuznetsova *et al.*, 2020). This function carries out a Satterthwaite approximation for degrees of freedom.

First, we built a 'full model' in which we tested the effect of season and intrinsic variables on FTM concentrations. This model included five fixed factors: sex, age, class (calves-atheel, weaned calves, adolescents, prime-aged elephants or elderly elephants), elephant origin (captive-born or wildcaught) and season (hot, cold or monsoon). To account for repeated measures on the same individuals, we included the ID of the elephants as a random factor. We also accounted for temporal and laboratory variation by including the year of sample collection and batch as random factors. When we faced singularity issues in the models, signalled by a warning from R, we removed the random factor(s) (year or batch) that had a variance near zero. In this first model, we also tested the interactions between the sex and age class of the elephants and between the sex and the season of the elephants to investigate whether FTM concentrations varied by age, and/or differently in males and females, and whether environmental conditions affected the FTM concentrations of males more than that of females.

From the first 'full model', we then determined a 'best model' to test the effect of FGM and H/L ratio. We selected this model using the second-order Akaike information criterion (AICc) given with the function 'dredge' of 'MuMIN' package (Burnham *et al.*, 2011; Barton, 2022). This best model kept only the variables explaining variation in FTM concentrations: the age class, sex and season, and the same random factors as the 'full model': elephants' ID, year, batch (see Table A6 in the appendix). To test the effect of stress variables, FGM and HL ratio, on FTM concentration variation, we separately added to the 'best model' one of these stress indicator variables at a time to determine its influence. We had to include these variables in separate models as the H/L

Table 1: Output of the linear mixed model, testing the effect of the origin of the elephants (captive-born or wild-caught), sex, age class and season on FTM concentrations, obtained with the summary of the model. As the interactions were not significant, the model was run again without the interactions, and we indicated only the *P* value given by the Type III analysis of variance. Values in bold indicate a significant *P* value.

Random effects	Variance		SD	
ID	0.001		0.029	
Batch	0.257		0.507	
Year	0.036		0.191	
	Estimate	SE	t	Р
Fixed effects				
Origin (wild-caught)	-2.691e-03	3.745e-02	-0.072	0.943
Season (hot)	2.152e-01	2.809e-02	7.661	2.55e-14***
Season (monsoon)	1.995e—01	2.600e-02	7.673	2.23e-14***
Age class (elderly elephants)	1.628e-01	6.397e-02	2.545	0.011*
Age class (weaned calves)	4.084e-02	5.624e-02	0.726	0.468
Age class (adolescents)	1.011e-01	5.658e-02	1.787	0.074
Age class (prime-aged elephants)	1.147e-01	5.955e-02	1.926	0.054
Sex (male)	6.850e-02	2.169e-02	3.158	0.002**
Sex * age class	NS	NS	NS	0.275
Sex * season	NS	NS	NS	0.299
Sample sizes				
Observations = 3163				
Individuals = 173				
AICc				
5173.4				

ratio was available for a smaller sample of animals and data points than FGM, since faecal samples were collected monthly versus once per season for blood samples. Thus, separate models were developed because including both variables in the same model would have unnecessarily limited the sample size of FGM.

Finally, we tested the effect of FTM concentration variation on the oxidative status of individuals. To do so, we used linear mixed models (package 'lme4', Bates *et al.*, 2015), adjusted with the maximum likelihood method, with FTM concentrations, age class, sex and season as fixed factors, and the ID of each individual, year of sample collection and plate number as random factors.

Results

FTM concentrations: association with sex, age class, origin and season

We found significant sex differences in variation in longitudinal FTM concentrations (Table 1), with males having 7.09% higher FTM concentration than females (Fig. 1). FTM concentrations also changed across life stages, increasing with age in both males and females (Table 1, age:sex interaction, P = 0.28). On average, elderly elephants had the highest FTM concentrations $(58.3 \pm 2.69 \text{ ng/g fae-}$ ces), followed by prime-aged elephants $(50.7 \pm 1.65 \text{ ng/g})$ faeces), adolescents $(49 \pm 1.31 \text{ ng/g faeces})$, calves-at-heel $(48.6 \pm 2.63 \text{ ng/g faeces})$ and weaned calves $(41.3 \pm 0.93 \text{ ng/g})$ faeces). Calves-at-heel and weaned calves did not significantly differ in their FTM concentrations (Table 1), whereas the differences between calves-at-heel and adolescent elephants, as well as between calves-at-heel and primeaged elephants were larger but non-significant (Table 1). FTM concentrations in elderly elephants were significantly higher than FTM concentrations in calves-at-heel (Table 1) and weaned calves ($\beta = 0.122 \pm 0.040$, t = 3.02, P = 0.003) elephants (Fig. 3). Moreover, both prime-aged elephants ($\beta = -0.074 \pm 0.033$, t = -2.26, P = 0.026) and adolescents ($\beta = -0.06 \pm 0.025$, t = -2.39, P = 0.017) had significantly higher FTM concentrations than weaned calves. We did not detect any effect of elephant origin (captive-born or wild-caught) on FTM concentrations (Table 1).



FIGURE 1: Comparison of FTM concentrations between male and female Asian elephants. The *Y* axis shows the back-transformed FTM concentrations with 95% confidence interval for males and females, corrected for the covariates in the final model in Table 1: season, age class and the origin of the elephants. The asterisk indicates a significant difference between FTM concentrations of males and females, obtained from linear mixed model.

In addition, we observed an effect of the environment, with significant seasonal variation in FTM concentrations (Table 1). Whilst elephants showed no significant difference in FTM concentrations between the hot and monsoon seasons ($\beta = 0.016 \pm 0.028$, t = 0.55, P = 0.58), their FTM concentrations were 24% higher in the hot season and 22% higher in monsoon than in the cold season (Table 1; Fig. 2). This seasonal variation in FTM concentrations was similar for males and females (season:sex interaction, P = 0.30, Table 1).

FTM concentrations: association with stress markers

We obtained different results for our two indicators of physiological stress: the H/L ratio and the concentration of FGM.

The H/L ratio ranged from 0.21 to 5.6, with an average value of 1.16 ± 0.04 . We found no significant link between the H/L ratio and FTM concentrations (Table 3).

By contrast, we found a strong positive relation between FTM concentrations and FGM concentrations (Table 2; Fig. 4). In our data, FGM concentrations ranged from 3.2 to 409.72 ng/g faeces, with an average value of 70.64 ± 0.57 ng/g faeces. For each one-unit increase in FGM, FTM concentrations increased by about 0.85%. Although the positive correlation between FTM concentrations and



FIGURE 2: Variation of FTM concentrations through life stages. The Y axis shows the back-transformed FTM concentrations with 95% confidence interval as a function of the age classes, corrected for the covariates in the final model in Table 1: sex, season and the status of the elephants. N indicates the number of individuals in each category. The results of the post hoc test are represented with letters above the graph. Two different letters for two modalities of the factor indicate that FTM concentrations are significantly different.

FGM concentrations seems driven by two extreme values of FGM in Fig. 4, the significance of the correlation remained similar after removing these data points from the analysis ($\beta = 0.009 \pm 4.1e - 4$, t = 22.25, P < 2.2e - 16).

FTM concentrations: association with oxidative status

Finally, we found no significant relationship between FTM concentrations and the concentration of ROMs (Table 4), products of oxidative stress or between FTM concentrations and the activity of SOD (Table 5), an enzyme with antioxidant activity.

Discussion

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Our results indicate that FTM concentration in the Asian elephant is a highly variable trait that responds to external and internal changes. We found that the concentrations of this androgen hormone differed between males and females, across different life stages and between seasons (which are known to affect health and mortality risks in the population) (Mumby *et al.*, 2013a, 2013b; dos Santos *et al.*, 2020). Specifically, FTM concentrations were overall higher in males than females, increased with life stages and were higher during

Table 2: Output of the linear mixed model testing the relation between FGM and FTM concentrations, obtained with the summary of the model. Values in bold indicate a significant *P* value.

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Random effects	Variance		SD	
ID	0.000		0.000	
Batch	0.461		0.679	
Year	0.063		0.250	
	Estimate	SE	t	Р
Fixed effects				
FGM	8.421e-03	3.861e-04	21.811	< 2e-16***
Sex (male)	7.819e-02	2.012e-02	3.887	1.04e-4***
Age class (elderly elephants)	1.757e—01	6.053e-02	2.902	0.004**
Age class (weaned calves)	7.301e-02	5.472e-02	1.334	0.182
Age class (adolescents)	1.344e-01	5.492e-02	2.447	0.014*
Age class (prime-aged elephants)	1.400e-01	5.577e-02	2.511	0.012*
Season (hot)	2.204e-01	2.740e-02	8.046	1.25e—15***
Season (monsoon)	2.108e-01	2.574e-02	8.190	3.85e-16***
Sample sizes				
Observations = 2964				
Individuals = 171				
AICc				
4486.7				

Table 3: Results of the linear mixed model, testing the relation between the H/L ratio and FTM concentrations, obtained with the summary of the model

Random effects	Variance		SD	
ID	0.000		0.000	
Batch	0.451		0.672	
	Estimate	SE	t	Р
Fixed effects				
H/L ratio	-0.002	0.047	-0.036	0.971
Sex (male)	0.129	0.064	2.006	0.046*
Age class (elderly elephants)	0.191	0.510	0.374	0.709
Age class (weaned calves)	0.106	0.499	0.213	0.832
Age class (adolescents)	0.322	0.501	0.642	0.521
Age class (prime-aged elephants)	0.231	0.504	0.458	0.648
Season (hot)	0.497	0.110	4.515	9.40e-06***
Season (monsoon)	0.249	0.331	0.753	0.453
Sample sizes				
Observations = 274				
Individuals = 89				
AICc				
436.4				



FIGURE 3: Variation of FTM concentrations through the three seasons in Myanmar. The Y axis shows back-transformed FTM concentrations with 95% confidence interval in the three different seasons, corrected for the covariates in the final model in Table 1: sex, age class and status of the elephants. *N* indicates the number of individuals in each category. The results of the *post hoc* test are represented with letters above the graph. Two different letters for two modalities of the factor indicate that FTM concentrations are significantly different.

monsoon and hot seasons compared to the cold season. Although we did not detect any link between H/L ratio and FTM concentrations, we found a strong positive correlation between FTM and FGM concentrations. Finally, we found no significant relation between FTM concentrations and the oxidative status of the individuals (i.e. ROM and SOD).

As expected, we observed significantly higher FTM concentrations in males than females, confirming the results of another study on Asian elephants (Rasmussen et al., 1984). This result is consistent with general observations in birds (Møller et al., 2005) and mammals, such as spotted hyenas (Glickman et al., 1987, 1992; Goymann et al., 2001), California ground squirrels (Holekamp and Talamantes, 1991), European moles (Whitworth et al., 1999), bonobos (Dittami et al., 2008) and dugongs (Burgess et al., 2012). However, the difference between sexes was not striking in our results, with males having 7.09% higher mean FTM concentrations than females (Fig. 1), likely due to the absence of males in musth in our data. In this study, we investigated the basal FTM levels in males and females. Males were sampled outside the musth, a periodic state in mature Asian and African bull elephants marked by elevated testosterone levels in blood, urine and faeces (Poole et al., 1984; Rasmussen et al., 1996; Sukumar, 2003; Brown et al., 2007; Kumar et al., 2014), associated with higher aggressiveness and temporal gland



FIGURE 4: FTM concentrations as a function of FGM concentrations. The Y axis shows the log-transformed FTM concentrations with 95% confidence intervals, corrected for the covariates season, age class and sex of the elephants as tested in the linear mixed model in Table 2. The line represents the predicted values of FTM concentrations, and N is the number of values/total number of individuals. Note that the association was even more pronounced when repeating the analysis without the two highest FGM values.

secretion (Rasmussen et al., 1984; Poole, 1987; Sukumar, 2003; Ganswindt et al., 2005). Our study indicates that outside of musth, FTM concentrations are surprisingly similar between the sexes. Unlike many other mammals living in seasonal environments, Asian elephants do not breed at a specific time of the year. Males can reproduce all year round and females have 14-16 weeks of reproductive cycle with an ovulation occurring in any season (Fowler and Mikota, 2006; Hildebrandt et al., 2011). This reproductive flexibility could explain the similar FTM concentrations between males and females. Additionally, social factors such as mating and competition might have influenced FTM concentrations, as it influences testosterone secretion in birds (Wingfield et al., 1990) and mammals (Taylor et al., 1989; McDonnell and Murray, 1995; Christensen et al., 2009), including Asian elephants (LaDue et al., 2023). We could not control for these variables in our study, but investigating their effects on androgen hormones could provide further insights.

Regarding the variation with age, we found an overall trend of increasing FTM concentrations across life stages in both sexes, with the highest concentrations observed in elderly elephants (Fig. 3). FTM concentrations were higher in adolescents, prime-aged elephants and elderly elephants compared to calves-at-heel and weaned calves, which are not yet sexually mature. Sexual maturity typically occurs at 14–

Table 4: Results of the linear mixed model, testing the effect of FTM concentrations on ROM, obtained with the summary of the model

Random effects	Variance		SD	
ID	1.526		1.235	
Plate	9.169		3.028	
Year	15.955		3.994	
	Estimate	SE	t	Р
Fixed effects				
FTM	0.003455	0.005808	0.595	0.55230
Sex (male)	1.045526	0.569493	1.836	0.07105
Age class (elderly elephants)	-0.035206	2.852814	-0.012	0.99016
Age class (weaned calves)	0.413090	2.524311	0.164	0.87011
Age class (adolescents)	0.470738	2.606332	0.181	0.85678
Age class (prime-aged elephants)	-0.179146	2.680152	-0.067	0.94675
Season (hot)	-2.624301	0.647511	-4.053	6.30e-05***
Season (monsoon)	-1.530216	0.571115	-2.679	0.00779**
Sample sizes				
Observations = 356				
Individuals = 81				
AICc				
2099.7				

TABLE 5: Results of the linear mixed model, testing the effect of FTM concentrations on SOD activity, obtained with the summary of the model

Random effects	Variance		SD	
ID	0.036		0.190	
Plate	0.142		0.377	
Year	0.010		0.098	
	Estimate	SE	t	Р
Fixed effects				
FTM	2.029e-04	5.616e-04	0.361	0.718
Sex (male)	-3.725e-02	6.623e-02	-0.562	0.576
Age class (elderly elephants)	5.864e-01	3.173e-01	1.848	0.065
Age class (weaned calves)	4.667e-01	2.841e-01	1.643	0.101
Age class (adolescents)	4.891e-01	2.934e-01	1.667	0.096
Age class (prime-aged elephants)	5.245e-01	3.007e-01	1.744	0.082
Season (hot)	-4.797e-02	6.322e-02	-0.759	0.449
Season (monsoon)	-8.734e-02	5.602e-02	-1.559	0.120
Sample sizes				
Observations = 438				
Individuals = 91				
AICc				
647.4				

15 years in males and females but can occur as early as 8-9 years (Shoshani and Eisenberg, 1982). When comparing FTM concentrations between immature (<14 years old) and mature individuals (see Table A5 in appendix), mature individuals showed significantly higher FTM levels, consistent with the findings of Brown et al. (2007), who reported a marked increase in testosterone levels in Asian elephants around 20 years of age. The surprisingly elevated FTM concentrations in calves-at-heel, which did not significantly differ from weaned calves, adolescents and prime-aged elephants, may result from hormone transfer during lactation. Hormonal transfer through mothers' milk has been demonstrated for glucocorticoids (Brummelte et al., 2010), which can influence offspring growth and behaviour, but such maternal transfer needs to be investigated for testosterone and in Asian elephants. Contrary to our hypothesis, elderly elephants (i.e. > 55 years old) exhibited the highest FTM concentrations. We did not observe a decrease in testosterone concentrations in older individuals, as reported by Thongtip et al. (2008) and Glaeser et al. (2022) in captive male Asian elephants. A potential explanation is the presence of a selective disappearance effect, where only high-quality individuals with higher FTM concentrations and longer lifespans remain in the oldest age class. Another explanation could be increased sexual activity at older ages. In both Asian and African elephants, males' social rank, which is size dependent, increases as they continue growing in size and weight with age (Poole, 1989; Lindeque and Jaarsveld, 1993; Lee and Moss, 1995; Sukumar, 2003; Chapman et al., 2016; Lalande et al., 2022). Larger, older African bulls are preferred sexual partners by females and have higher reproductive success (Sukumar, 2003). Moreover, while previous studies have focused primarily on males, our findings were consistent between males and females, with no significant interaction between sex and age class. In female Asian elephants, testosterone levels positively correlate with progesterone concentrations during the oestrous cycle and increase during the luteal phase (Taya et al., 1991). This suggests the production of testosterone by the ovaries in this genus (Taya et al., 1991), a phenomenon well-known in humans (e.g. Kirschner and Wayne Bardin, 1972; Burger, 2002). Hence, testosterone levels may also rise in females as they reach sexual maturity. Furthermore, Asian elephants remain sexually active and can reproduce into advanced ages, with reproduction beyond 60 years reported in females (Sukumar, 2003; Lahdenperä et al., 2014). The higher sexual activity of older elephants may explain the observed increase in FTM concentrations, but further research is needed, especially in females, to better understand this pattern.

As expected, we evidenced strong seasonal variation in FTM concentrations. The climate in Myanmar is defined by three distinct seasons. The hot season is characterized by low humidity, high temperatures and low food availability. In contrast, monsoon is a rainy season with significant rainfall, high temperatures, higher food availability and easier access to water. Finally, the cold season is marked by low temperatures, lower precipitation and better food availability than the hot

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season, especially at the beginning of the season. Previous studies on MTE elephants found that these seasonal changes influence the elephants' health, survival and stress markers (FGM and H/L ratio) (Mumby et al., 2013a; dos Santos et al., 2020; Ukonaho et al., 2023). Specifically, stress markers were higher during the cold season (Ukonaho et al., 2023) and survival rates were higher during monsoon (Mumby et al., 2013a). In this study, we observed higher FTM concentrations during monsoon and the hot season than the cold season for both sexes. Glucocorticoids might have suppressed testosterone secretion by inhibiting the HPG axis during the cold season. However, we cannot confirm this hypothesis here, as we found a positive correlation between biomarkers of stress and FTM concentrations. In Thailand, captive male Asian elephants also exhibit higher testosterone concentrations during monsoon (Thongtip et al., 2008; Norkaew et al., 2019). However, contrary to our results, they show higher testosterone concentrations in the cold than in the hot season. The seasonal variation in FTM concentrations is likely influenced by fluctuations in food availability, as suggested by Cooper et al. (1990). Testosterone secretion is tightly linked to diet, food intake and nutritional status in male vertebrates (Wilson et al., 1979; Wingfield, 1988; Pérez-Rodríguez et al., 2006; Ruiz et al., 2010). Monsoon is an energetically favourable season with better food availability, providing elephants with a higher-quality diet, which could explain the higher FTM concentrations observed during this season (Thongtip et al., 2008; Norkaew et al., 2019). In addition, when male captive Asian elephants are fed ad libitum, FTM concentration correlates positively with temperature and humidity (de Andrés et al., 2021), suggesting that these seasonal factors also affect testosterone secretion. For female captive Asian elephants, body condition is better during monsoon, the luteal phase and oestrous cycle are longer and peaks of progesterone levels are higher (Yang et al., 2023), which correlates positively with testosterone concentrations (Taya et al., 1991). Thus, the seasonal variation in food availability may influence body condition in elephants and thereby affecting testosterone secretion by the testes in males and the ovaries in females. Finally, the higher FTM concentrations observed during both monsoon and the hot season might be related to the reproductive activity of the elephants. Musth in male Asian elephants is more observed during monsoon (Kahl and Armstrong, 2002), and a peak of conceptions in MTE elephants occurs during the hot season (Mumby et al., 2013b, 2015), reflecting a higher sexual activity during these seasons. Hence, FTM concentrations in elephants are influenced by seasonal environmental variations, which might be underpinned by fluctuating food availability and increased sexual activity.

Our results on the relationship between stress measures and FTM concentrations are contradictory. We found no association between FTM concentrations and the H/L ratio. Although this result differs from our predictions, a previous study on the house sparrow found that an experimental elevation of testosterone concentrations with testosterone implants did not change the H/L ratio (Buchanan

et al., 2003), suggesting an absence of association between testosterone secretion and stress response. This analysis was performed on a smaller sample size, which may have also reduced statistical power in our study. However, we found a particularly strong relationship between FTM and FGM concentrations using a larger sample size. While stress and glucocorticoids are typically thought to suppress the HPG axis and inhibit reproductive functions (e.g. Saketos et al., 1993; Wingfield and Sapolsky, 2003; Narayan and Parisella, 2017), the positive correlation observed here between and rogen and adrenal hormones aligns with findings from other studies on boar (Liptrap and Raeside, 1975, 1978; Borg et al., 1991), pygmy goats (Howland et al., 1985), bison bulls (Mooring et al., 2006) and Asian elephant bulls during musth (Rasmussen et al., 1984; Brown et al., 2007; Yon et al., 2007b, 2007a, 2008; Kumar et al., 2014) (but see de Andrés et al., 2021). The correlation between FGM and FTM may be explained by the release of androgens from the Zona reticularis of the adrenal gland by stimulation of the HPA axis. In addition to glucocorticoids, the HPA axis is involved in the production of adrenal androgens such as dehydroepiandrosterone (Antoniou-Tsigkos et al., 2019; Dutt et al., 2023), which can then be converted into other androgens, including testosterone. Thus, some testosterone may be produced alongside glucocorticoids, potentially explaining the positive correlation observed here. This correlation might also be due to indirect processes. Organisms can develop mechanisms to maintain reproduction under stress, reviewed in Wingfield and Sapolsky (2003). The reproductive system could be resistant to glucocorticoids, especially with chronic stress, and/or the stimulation of the HPG axis, and testosterone secretion could increase to compensate for the inhibitory effects of glucocorticoids to enable reproduction under stress (Place and Kenagy, 2000; Lidgard et al., 2008). Alternatively, stress can occur with reproduction. An elevation of glucocorticoid concentrations is observed during the breeding period in amphibians, reptiles (Moore and Jessop, 2003), Australian marsupials (Bradley et al., 1980; McDonald et al., 1981) and sexual maturation in the Pacific Sockeye salmon (Donaldson and Fagerlund, 1968, 1970). Similarly, sexual signalling, courtships, vocalization and aggressive behaviours require high energetic expenditure, leading to increased corticosterone secretion (Moore and Jessop, 2003). In this study, we lack behavioural data to parallel the physiological markers of stress (Seltmann et al., 2020). Further investigations including behavioural observations are needed to better understand the association between markers of stress and FTM concentrations. Finally, both FGM and FTM concentrations reflect hormonal levels in elephants over approximately 24 hours, while the H/L ratio, influenced by blood cell turnover (30-40 days), captures stress on a different timescale. The contradictory results we observed between these markers of stress may be explained by the differing temporal nature of these physiological measurements.

Regarding the indicators of oxidative stress and the molecular consequences of variations in FTM concentrations, we found no significant relation between FTM concentrations and either ROMs or antioxidant enzyme activity (SOD). The CV from these oxidative stress measurements suggests there is some noise in the measurements, which might explain this absence of a relation. Our results align with Isaksson et al. (2011), who reported similar findings in a natural population of White's skinks. Nonetheless, in the literature, studies examining the relation between testosterone and oxidative stress have produced conflicting results. While elevated testosterone concentrations have been shown to increase oxidative stress, potentially leading to important fitness costs (Chainy et al., 1997; Zhu et al., 1997; Pansarasa et al., 2002; Aydilek et al., 2004; Alonso-Alvarez et al., 2007), other studies suggest that testosterone may have antioxidant properties (Ahlbom et al., 2001; Tam et al., 2003; Guzmán et al., 2005). Hence, we still know little about the effect of testosterone variation on the oxidative status of individuals, and further research is needed to clarify this relation. Finally, a recent study on the same elephant population in Myanmar found no seasonal variation in ROM and SOD levels (Ukonaho et al., 2023). While we observed a significant effect of season on ROM in our results (Table 4), this likely reflects an artefact of the reduced sample size when oxidative stress markers were merged with FTM data. Thus, oxidative stress levels in these elephants appear stable despite seasonal changes and fluctuations in FTM concentrations.

To conclude, our results confirm that FTM concentrations fluctuate with age, sex, seasonal variation and stress levels. Understanding both the intrinsic and extrinsic factors that influence FTM concentration is crucial for comprehending how this hormone shapes individuals' traits in a changing environment, leading to a diversity of endocrinological phenotypes within and across populations. Listed as endangered on the IUCN red list (Williams et al., 2019), the Asian elephant population is in decline, largely due to habitat loss, fragmentation and vital rates that do not support population growth. A quarter of the remaining Asian elephants are now in captivity, yet these populations also struggle with low birth rates and high infant mortality (Jackson et al., 2019). By identifying the factors influencing FTM concentrations, our study offers critical insights to enhance conservation efforts for this endangered species. As an androgen hormone, testosterone plays a key role in regulating reproductive physiology and behaviour, directly affecting individual fitness. Knowledge of its determinants is therefore essential for optimizing fertility and reproductive success in captive populations. Additionally, understanding FTM variation in semi-captive populations provides important insights into the reproductive health of wild populations and their adaptation to environmental changes. Ultimately, this knowledge is essential for Asian elephants' conservation, to improve environmental management and implement effective conservation strategies for this endangered species.

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Author Contributions

H.M.: Data curation, data analyses, visualization, writing (first draft). V.B.: data analyses, review of the manuscript. D.S.: Data collection, laboratory analyses, review of the manuscript. S.U.: Laboratory analyses, review of the manuscript. L.Y.: Review of the manuscript. M.B.: Data analyses, review of the manuscript. U.K.N.: Data collection, data curation, review of the manuscript. V.L.: Conceptualization, supervision, review of the manuscript. S.R.: Conceptualization, supervision, review of the manuscript.

Conflicts of interest

The authors have no conflicts of interest to declare.

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Data Availability

Data available on request.

Supplementary Materials

Supplementary material is available at Conservation Physiology online.

References

- Aakvaag A, Bentdal Ø, Quigstad K, Walstad P, Rønningen H, Fonnum F (1978) Testosterone and testosterone binding globulin (TeBG) in young men during prolonged stress. *Int J Androl* 1: 22–31. https:// doi.org/10.1111/j.1365-2605.1978.tb00573.x.
- Ahlbom E, Prins GS, Ceccatelli S (2001) Testosterone protects cerebellar granule cells from oxidative stress-induced cell death through a receptor mediated mechanism. *Brain Res* 892: 255–262. https://doi. org/10.1016/S0006-8993(00)03155-3.

.....

- Alonso-Alvarez C, Bertrand S, Faivre B, Chastel O, Sorci G (2007) Testosterone and oxidative stress: the oxidation handicap hypothesis. *Proc R Soc B Biol Sci* 274: 819–825. https://doi.org/10.1098/ rspb.2006.3764.
- de Andrés PJ, Cáceres S, Crespo B, Silván G, Illera JC (2021) Non-invasive determination of annual fecal cortisol, androstenedione, and testos-terone variations in a herd of male Asian elephants (*Elephas maximus*) and their relation to some climatic variables. *Animals* 11: 2723. https://doi.org/10.3390/ani11092723.
- Antoniou-Tsigkos A, Zapanti E, Ghizzoni L, Mastorakos G *et al.* (2019) Adrenal Androgens. In KR Feingold, B Anawalt, MR Blackman *et al.*, eds, *Endotext* [Internet]. MDText.com, Inc, South Dartmouth (MA).
- Astheimer LB, Buttemer WA, Wingfield JC (1995) Seasonal and acute changes in adrenocortical responsiveness in an Arctic-breeding bird. *Horm Behav* 29: 442–457. https://doi.org/10.1006/hbeh.1995.1276.
- Aydilek N, Aksakal M, Karakılçık AZ (2004) Effects of testosterone and vitamin E on the antioxidant system in rabbit testis. *Andrologia* 36: 277–281. https://doi.org/10.1111/j.1439-0272.2004.00618.x.

Barton K (2022) MuMIn: multi-model inference.

- Bates D, Mächler M, Bolker B, Walker S (2015) Fitting linear mixed-effects models using Ime4. *J Stat Softw* 67: 1–48. https://doi.org/10.18637/jss.v067.i01.
- Beehner JC, Gesquiere L, Seyfarth RM, Cheney DL, Alberts SC, Altmann J (2009) Testosterone related to age and life-history stages in male baboons and geladas. *Horm Behav* 56: 472–480. https://doi. org/10.1016/j.yhbeh.2009.08.005.
- Beehner JC, Phillips-Conroy JE, Whitten PL (2005) Female testosterone, dominance rank, and aggression in an Ethiopian population of hybrid baboons. *Am J Primatol* 67: 101–119. https://doi.org/10.1002/ajp.20172.
- Borg KE, Esbenshade KL, Johnson BH (1991) Cortisol, growth hormone, and testosterone concentrations during mating behavior in the bull and boar1. *J Anim Sci* 69: 3230–3240. https://doi.org/10.2527/ 1991.6983230x.
- Bradley AJ, McDonald IR, Lee AK (1980) Stress and mortality in a small marsupial (*Antechinus stuartii*, Macleay). *Gen Comp Endocrinol* 40: 188–200. https://doi.org/10.1016/0016-6480(80)90122-7.
- Briga M, Jimeno B, Verhulst S (2019) Coupling lifespan and aging? The age at onset of body mass decline associates positively with sex-specific lifespan but negatively with environment-specific lifespan. *Exp Gerontol* 119: 111–119. https://doi.org/10.1016/j.exger.2019. 01.030.
- Briga M, Verhulst S (2021) Mosaic metabolic ageing: basal and standard metabolic rates age in opposite directions and independent of environmental quality, sex and life span in a passerine. *Funct Ecol* 35: 1055–1068. https://doi.org/10.1111/1365-2435.13785.
- Brown JL, Somerville M, Riddle HS, Keele M, Duer CK, Freeman EW (2007) Comparative endocrinology of testicular, adrenal and thyroid function in captive Asian and African elephant bulls. *Gen Comp Endocrinol* 151: 153–162. https://doi.org/10.1016/j.ygcen.2007.01.006.

- Brummelte S, Schmidt KL, Taves MD, Soma KK, Galea LAM (2010) Elevated corticosterone levels in stomach milk, serum, and brain of male and female offspring after maternal corticosterone treatment in the rat. *Dev Neurobiol* 70: 714–725. https://doi.org/10.1002/dneu.20805.
- Buchanan KL, Evans MR, Goldsmith AR (2003) Testosterone, dominance signalling and immunosuppression in the house sparrow, Passer domesticus. *Behav Ecol Sociobiol* 55: 50–59. https://doi.org/10.1007/ s00265-003-0682-4.
- Burger HG (2002) Androgen production in women. *Fertil Steril* 77: 3–5. https://doi.org/10.1016/S0015-0282(02)02985-0.
- Burgess EA, Lanyon JM, Keeley T (2012) Testosterone and tusks: maturation and seasonal reproductive patterns of live, free-ranging male dugongs (Dugong dugon) in a subtropical population. *Reproduction* 143: 683–697. https://doi.org/10.1530/REP-11-0434.
- Burnham KP, Anderson DR, Huyvaert KP (2011) AIC model selection and multimodel inference in behavioral ecology: some background, observations, and comparisons. *Behav Ecol Sociobiol* 65: 23–35. https://doi.org/10.1007/s00265-010-1029-6.
- Cai Z, Li H (2020) An updated review: androgens and cognitive impairment in older men. *Front Endocrinol* 11. https://doi.org/10.3389/ fendo.2020.586909.
- Chainy GBN, Samantaray S, Samanta L (1997) Testosterone-induced changes in testicular antioxidant system. *Andrologia* 29: 343–349. https://doi.org/10.1111/j.1439-0272.1997.tb00328.x.
- Chapman SN, Mumby HS, Crawley JAH, Mar KU, Htut W, Soe AT, Aung HH, Lummaa V (2016) How big is it really? Assessing the efficacy of indirect estimates of body size in Asian elephants. *PLoS One* 11: e0150533. https://doi.org/10.1371/journal.pone.0150533.
- Christensen BW, Troedsson MHT, Young LJ, Oliva M, Penfold LM (2009) Effects of sociosexual environment on serum testosterone in captive male African rhinoceros. *Theriogenology* 71: 1105–1111. https://doi. org/10.1016/j.theriogenology.2008.12.003.
- Cooper KA, Harder JD, Clawson DH, Fredrick DL, Lodge GA, Peachey HC, Spellmire TJ, Winstel DP (1990) Serum testosterone and musth in captive male African and Asian elephants. *Zoo Biol* 9: 297–306. https://doi.org/10.1002/zoo.1430090405.
- Costantini D (2016) Oxidative stress ecology and the d-ROMs test: facts, misfacts and an appraisal of a decade's work. *Behav Ecol Sociobiol* 70: 809–820. https://doi.org/10.1007/s00265-016-2091-5.
- Costantini D, Dell'Omo G (2006) Effects of T-cell-mediated immune response on avian oxidative stress. *Comp Biochem Physiol A Mol Integr Physiol* 145: 137–142. https://doi.org/10.1016/j.cbpa.2006.06.002.
- Crawford BA, Harewood WJ, Handelsman DJ (1997) Growth and hormone characteristics of pubertal development in the hamadryas baboon. *J Med Primatol* 26: 153–163. https://doi.org/10.1111/ j.1600-0684.1997.tb00047.x.
- Crawley JAH, Lahdenperä M, Min Oo Z, Htut W, Nandar H, Lummaa V (2020) Taming age mortality in semi-captive Asian elephants. *Sci Rep* 10: 1889. https://doi.org/10.1038/s41598-020-58590-7.

- Crawley JAH, Liehrmann O, Franco dos Santos DJ, Brown J, Nyein UK, Aung HH, Htut W, Oo ZM, Seltmann MW, Webb JL *et al.* (2021) Influence of handler relationships and experience on health parameters, glucocorticoid responses and behaviour of semi-captive Asian elephants. *Conserv Physiol* 9: coaa116. https://doi.org/10.1093/conphys/coaa116.
- Cruz-Topete D, Dominic P, Stokes KY (2020) Uncovering sex-specific mechanisms of action of testosterone and redox balance. *Redox Biol* 31: 101490. https://doi.org/10.1016/j.redox.2020.101490.
- Davis AK, Maney DL (2018) The use of glucocorticoid hormones or leucocyte profiles to measure stress in vertebrates: What's the difference? *Methods Ecol Evol* 9: 1556–1568. https://doi.org/10.1111/2041-210X.13020.
- Dittami J, Katina S, Möstl E, Eriksson J, Machatschke IH, Hohmann G (2008) Urinary androgens and cortisol metabolites in field-sampled bonobos (*Pan paniscus*). *Gen Comp Endocrinol* 155: 552–557. https:// doi.org/10.1016/j.ygcen.2007.08.009.
- Donaldson EM, Fagerlund UHM (1968) Changes in the cortisol dynamics of sockeye salmon (*Oncorhynchus nerka*) resulting from secual maturation. *Gen Comp Endocrinol* 11: 552–561. https://doi.org/10.1016/ 0016-6480(68)90071-3.
- Donaldson EM, Fagerlund UHM (1970) Effect of sexual maturation and gonadectomy at sexual maturity on cortisol secretion rate in sockeye salmon (*Oncorhynchus nerka*). J Fish Res Board Can 27: 2287–2296. https://doi.org/10.1139/f70-256.
- Dufty AM (1989) Testosterone and survival: a cost of aggressiveness? *Horm Behav* 23: 185–193. https://doi.org/10.1016/0018-506 X(89)90059-7.
- Dutt M, Wehrle CJ, Jialal I (2023) Physiology, Adrenal Gland. In *StatPearls*. StatPearls Publishing, Treasure Island (FL)
- Finnegan D (2015) referenceIntervals: Reference Intervals. https://cran.rproject.org/web/packages/referenceIntervals/index.html.
- Fowler ME, Mikota SK (2006) *Biology, Medicine, and Surgery of Elephants*. Wiley-Blackwell, Ames (IA), p. 565.
- Ganswindt A, Rasmussen HB, Heistermann M, Hodges JK (2005) The sexually active states of free-ranging male African elephants (*Loxodonta africana*): defining musth and non-musth using endocrinology, physical signals, and behavior. *Horm Behav* 47: 83–91. https://doi.org/10.1016/j.yhbeh.2004.09.002.
- Glaeser SS, Edwards KL, Paris S, Scarlata C, Lee B, Wielebnowski N, Finnell S, Somgird C, Brown JL (2022) Characterization of longitudinal testosterone, cortisol, and musth in male Asian elephants (*Elephas maximus*), effects of aging, and adrenal responses to social changes and health events. *Animals* 12: 1332. https://doi. org/10.3390/ani12101332.
- Glaeser SS, Shepherdson D, Lewis K, Prado N, Brown JL, Lee B, Wielebnowski N (2021) Supporting zoo Asian elephant (*Elephas maximus*) welfare and herd dynamics with a more complex and expanded habitat. *Animals* 11: 2566. https://doi.org/10.3390/ ani11092566.

- Glickman SE, Frank LG, Davidson JM, Smith ER, Siiteri PK (1987) Androstenedione may organize or activate sex-reversed traits in female spotted hyenas. *Proc Natl Acad Sci* 84: 3444–3447. https://doi. org/10.1073/pnas.84.10.3444.
- Glickman SE, Frank LG, Pavgi S, Licht P (1992) Hormonal correlates of "masculinization" in female spotted hyaenas (*Crocuta crocuta*). 1. Infancy to sexual maturity. *Reproduction* 95: 451–462. https://doi. org/10.1530/jrf.0.0950451.
- Goymann W, East ML, Hofer H (2001) Androgens and the role of female "hyperaggressiveness" in spotted hyenas (*Crocuta crocuta*). Horm Behav 39: 83–92. https://doi.org/10.1006/hbeh.2000.1634.
- Gray GD, Smith ER, Damassa DA, Ehrenkranz JRL, Davidson JM (2004) Neuroendocrine mechanisms mediating the suppression of circulating testosterone levels associated with chronic stress in male rats. *Neuroendocrinology* 25: 247–256. https://doi.org/10.1159/ 000122746.
- Gross WB, Siegel HS (1983) Evaluation of the heterophil/lymphocyte ratio as a measure of stress in chickens. *Avian Dis* 27: 972–979. https://doi.org/10.2307/1590198.
- Gündoğan M (2007) Seasonal variation in serum testosterone, T3 and andrological parameters of two Turkish sheep breeds. *Small Rumin Res* 67: 312–316. https://doi.org/10.1016/j.smallrumres.2005.11.005.
- Guzmán DC, Mejía GB, Vázquez IE, García EH, Angel DS d, Olguín HJ (2005) Effect of testosterone and steroids homologues on indolamines and lipid peroxidation in rat brain. *J Steroid Biochem Mol Biol* 94: 369–373. https://doi.org/10.1016/j.jsbmb.2004.11.006.
- Halliwell B (1994) Free radicals and antioxidants: a personal view. *Nutr Rev* 52: 253–265. https://doi.org/10.1111/j.1753-4887.1994. tb01453.x.
- Hegner RE, Wingfield JC (1987) Effects of experimental manipulation of testosterone levels on parental investment and breeding success in male house sparrows. *Auk* 104: 462–469. https://doi. org/10.2307/4087545.
- Hildebrandt TB, Lueders I, Hermes R, Goeritz F, Saragusty J (2011) Reproductive cycle of the elephant. *Anim Reprod Sci* 124: 176–183. https:// doi.org/10.1016/j.anireprosci.2010.08.027.
- Holekamp KE, Talamantes F (1991) Seasonal variation in circulating testosterone and oestrogens of wild-caught California ground squirrels (Spermophilus beecheyi). *J Reprod Fertil* 93: 415–425. https://doi. org/10.1530/jrf.0.0930415.
- Howland BE, Sanford LM, Palmer WM (1985) Changes in serum levels of LH, FSH, prolactin, testosterone, and cortisol associated with season and mating in male pygmy goats. *J Androl* 6: 89–96. https://doi. org/10.1002/j.1939-4640.1985.tb00822.x.
- Isaksson C, While GM, McEvoy J, van de Crommenacker J, Olsson M, Groothuis TGG, Komdeur J, Wapstra E (2011) Aggression, but not testosterone, is associated to oxidative status in a free-living vertebrate. *Behaviour* 148: 713–731. https://doi.org/10.1163/000579511 X574204.

- Jackson J, Childs DZ, Mar KU, Htut W, Lummaa V (2019) Long-term trends in wild-capture and population dynamics point to an uncertain future for captive elephants. *Proc R Soc B Biol Sci* 286: 20182810. https://doi.org/10.1098/rspb.2018.2810.
- Kahl MP, Armstrong BD (2002) Visual displays of wild African elephants during musth. 66: 159–172.
- Keay JM, Singh J, Gaunt MC, Kaur T (2006) Fecal glucocorticoids and their metabolites AS indicators of stress IN various mammalian species: a literature review. J Zoo Wildl Med 37: 234–244. https://doi. org/10.1638/05-050.1.
- Kempenaers B, Peters A, Foerster K (2008) Sources of individual variation in plasma testosterone levels. *Philos Trans R Soc B* 363: 1711–1723. https://doi.org/10.1098/rstb.2007.0001.
- Ketterson ED, Nolan V Jr, Cawthorn MJ, Parker PG, Ziegenfus C (1996) Phenotypic engineering: using hormones to explore the mechanistic and functional bases of phenotypic variation in nature. *Ibis* 138: 70–86. https://doi.org/10.1111/j.1474-919X.1996.tb04314.x.
- Ketterson ED, Val NJ (1999) Adaptation, exaptation, and constraint: a hormonal perspective. Am Nat 154: S4–S25. https://doi.org/10.1086/ 303280.
- Kirschner MA, Wayne Bardin C (1972) Androgen production and metabolism in normal and virilized women. *Metabolism* 21: 667–688. https://doi.org/10.1016/0026-0495(72)90090-X.
- Kreuz MLE, Rose RM, Jennings CJR (1972) Suppression of plasma testosterone levels and psychological stress: a longitudinal study of young men in officer candidate school. Arch Gen Psychiatry 26: 479–482. https://doi.org/10.1001/archpsyc.1972.01750230089017.
- Kumar V, Palugulla Reddy V, Kokkiligadda A, Shivaji S, Umapathy G (2014) Non-invasive assessment of reproductive status and stress in captive Asian elephants in three south Indian zoos. *Gen Comp Endocrinol* 201: 37–44. https://doi.org/10.1016/j.ygcen.2014. 03.024.
- Kuznetsova A, Brockhoff PB, Christensen RHB, Jensen SP (2020) ImerTest: tests in linear mixed effects models.
- LaDue CA, Hunt KE, Kiso WK, Freeman EW (2023) Hormonal variation and temporal dynamics of musth in Asian elephants (*Elephas maximus*) are associated with age, body condition and the social environment. *Conserv Physiol* 11: coad019. https://doi.org/10.1093/conphys/ coad019.
- LaDue CA, Hunt KE, Samaraweera MGSM, Vandercone RPG, Kiso WK, Freeman EW (2022) Physical and behavioral indicators associated with hormonal changes during musth in zoo-housed and freeranging Asian elephants (*Elephas maximus*). *Theriogenology Wild* 1: 100011. https://doi.org/10.1016/j.therwi.2022.100011.
- Lahdenperä M, Mar KU, Courtiol A, Lummaa V (2018) Differences in age-specific mortality between wild-caught and captive-born Asian elephants. *Nat Commun* 9: 3023. https://doi.org/10.1038/s41467-018-05515-8.

Lahdenperä M, Mar KU, Lummaa V (2014) Reproductive cessation and post-reproductive lifespan in Asian elephants and preindustrial humans. *Front Zool* 11:54. https://doi.org/10.1186/s12983-014-0054-0.

.....

- Lalande LD, Lummaa V, Aung HH, Htut W, Nyein UK, Berger V, Briga M (2022) Sex-specific body mass ageing trajectories in adult Asian elephants. *J Evol Biol* 35: 752–762. https://doi.org/10.1111/jeb.14008.
- Lee PC, Moss CJ (1995) Statural growth in known-age African elephants (*Loxodonta africana*). J Zool 236: 29–41. https://doi.org/10.1111/j.1469-7998.1995.tb01782.x.
- Lidgard DC, Boness DJ, Bowen WD, McMillan JI (2008) The implications of stress on male mating behavior and success in a sexually dimorphic polygynous mammal, the grey seal. *Horm Behav* 53: 241–248. https://doi.org/10.1016/j.yhbeh.2007.10.003.
- Lindeque M, Jaarsveld AS (1993) Post-natal growth of elephants *Lox-odonta africana* in Etosha National Park, Namibia. *J Zool* 229: 319–330. https://doi.org/10.1111/j.1469-7998.1993.tb02639.x.
- Liptrap RM, Raeside JI (1975) Increase IN plasma testosterone concentration after injection of adrenocorticotrophin into the boar. *J Endocrinol* 66: 123–131. https://doi.org/10.1677/joe.0.0660123.
- Liptrap RM, Raeside JI (1978) A relationship between plasma concentrations of testosterone and corticosteroids during sexual and aggressive behaviour IN the boar. *J Endocrinol* 76: 75–85. https://doi. org/10.1677/joe.0.0760075.
- Mar KU, Lahdenperä M, Lummaa V (2012) Causes and correlates of calf mortality in captive Asian elephants (*Elephas maximus*). *PLoS One* 7: e32335. https://doi.org/10.1371/journal.pone.0032335.
- Marler CA, Moore MC (1988) Evolutionary costs of aggression revealed by testosterone manipulations in free-living male lizards. *Behav Ecol Sociobiol* 23: 21–26. https://doi.org/10.1007/BF00303053.
- Martin DE, Swenson RB, Collins DC (1977) Correlation of serum testosterone levels with age in male chimpanzees. *Steroids* 29: 471–481. https://doi.org/10.1016/0039-128X(77)90067-8.
- Matsumoto K, Takeyasu K, Mizutani S, Hamanaka Y, Uozumi T (1970) Plasma testosterone levels following surgical stress IN male patients. *Eur J Endocrinol* 65: 11–17. https://doi.org/10.1530/acta.0.0650011.
- McDonald IR, Lee AK, Bradley AJ, Than KA (1981) Endocrine changes in dasyurid marsupials with differing mortality patterns. *Gen Comp Endocrinol* 44: 292–301. https://doi.org/10.1016/0016-6480 (81)90004-6.
- McDonnell SM, Murray SC (1995) Bachelor and harem stallion behavior and endocrinology. *Biol Reprod* 52: 577–590. https://doi.org/ 10.1093/biolreprod/52.monograph_series1.577.
- McFarlane JM, Curtis SE (1989) Multiple concurrent stressors in chicks: 3. Effects on plasma corticosterone and the heterophil:lymphocyte ratio. *Poult Sci* 68: 522–527. https://doi.org/10.3382/ps.0680522.
- McGlothlin JW, Jawor JM, Ketterson ED (2007) Natural variation in a testosterone-mediated trade-off between mating effort and

.....

parental effort. *Am Nat* 170: 864–875. https://doi.org/10.1086/ 522838.

- Meydan S, Kus I, Tas U, Ogeturk M, Sancakdar E, Dabak DO, Zararsız I, Sarsılmaz M (2010) Effects of testosterone on orchiectomy-induced oxidative damage in the rat hippocampus. *J Chem Neuroanat* 40: 281–285. https://doi.org/10.1016/j.jchemneu.2010.07.006.
- Mills SC, Grapputo A, Jokinen I, Koskela E, Mappes T, Oksanen TA, Poikonen T (2009) Testosterone-mediated effects on fitness-related phenotypic traits and fitness. *Am Nat* 173: 475–487. https://doi. org/10.1086/597222.
- Moe RO, Bakken M (1996) Effect of repeated blood sampling on plasma concentrations of cortisol and testosterone and on leucocyte number in silver fox vixens (Vulpes Vulpes). *Acta Agric Scand Sect Anim Sci* 46: 111–116.
- Møller AP, Garamszegi LZ, Gil D, Hurtrez-Boussès S, Eens M (2005) Correlated evolution of male and female testosterone profiles in birds and its consequences. *Behav Ecol Sociobiol* 58: 534–544. https://doi. org/10.1007/s00265-005-0962-2.
- Moore IT, Jessop TS (2003) Stress, reproduction, and adrenocortical modulation in amphibians and reptiles. *Horm Behav* 43: 39–47. https://doi.org/10.1016/S0018-506X(02)00038-7.
- Moore MC, Thompson CW, Marler CA (1991) Reciprocal changes in corticosterone and testosterone levels following acute and chronic handling stress in the tree lizard, *Urosaurus ornatus*. *Gen Comp Endocrinol* 81: 217–226. https://doi.org/10.1016/0016-6480(91)90006-R.
- Mooring MS, Patton ML, Lance VA, Hall BM, Schaad EW, Fetter GA, Fortin SS, McPeak KM (2006) Glucocorticoids of bison bulls in relation to social status. *Horm Behav* 49: 369–375. https://doi.org/10.1016/j. yhbeh.2005.08.008.
- Mumby HS, Courtiol A, Mar KU, Lummaa V (2013a) Climatic variation and age-specific survival in Asian elephants from Myanmar. *Ecology* 94: 1131–1141. https://doi.org/10.1890/12-0834.1.
- Mumby HS, Courtiol A, Mar KU, Lummaa V (2013b) Birth seasonality and calf mortality in a large population of Asian elephants. *Ecol Evol* 3: 3794–3803. https://doi.org/10.1002/ece3.746.
- Mumby HS, Mar KU, Thitaram C, Courtiol A, Towiboon P, Min-Oo Z, Htut-Aung Y, Brown JL, Lummaa V (2015) Stress and body condition are associated with climate and demography in Asian elephants. *Conserv Physiol* 3: cov030. https://doi.org/10.1093/conphys/cov030.
- Narayan E, Parisella S (2017) Influences of the stress endocrine system on the reproductive endocrine axis in sheep (*Ovis aries*). *Ital J Anim Sci* 16: 640–651. https://doi.org/10.1080/1828051X.2017.1321972.
- Negro SS, Caudron AK, Dubois M, Delahaut P, Gemmell NJ (2010) Correlation between male social status, testosterone levels, and parasitism in a dimorphic polygynous mammal. *PLoS One* 5: e12507. https://doi. org/10.1371/journal.pone.0012507.
- Norkaew T, Brown JL, Bansiddhi P, Somgird C, Thitaram C, Punyapornwithaya V, Punturee K, Vongchan P, Somboon N, Khonmee J (2019) Influence of season, tourist activities and camp

management on body condition, testicular and adrenal steroids, lipid profiles, and metabolic status in captive Asian elephant bulls in Thailand. *PLoS One* 14: e0210537. https://doi.org/10.1371/journal. pone.0210537.

- O'Neal DM, Reichard DG, Pavilis K, Ketterson ED (2008) Experimentallyelevated testosterone, female parental care, and reproductive success in a songbird, the dark-eyed junco (*Junco hyemalis*). *Horm Behav* 54: 571–578. https://doi.org/10.1016/j.yhbeh.2008.05.017.
- Oring LW, Fivizzani AJ, El Halawani ME (1989) Testosterone-induced inhibition of incubation in the spotted sandpiper (*Actitis mecularia*). *Horm Behav* 23: 412–423. https://doi.org/10.1016/0018-506 X(89)90053-6.
- Orlando EF, Binczik GA, Thomas P, Guillette LJ (2003) Reproductive seasonality of the male Florida gar, *Lepisosteus platyrhincus. Gen Comp Endocrinol* 131: 365–371. https://doi.org/10.1016/S0016-6480 (03)00036-4.
- Padmanabhan V, Manikkam M, Recabarren S, Foster D (2006) Prenatal testosterone excess programs reproductive and metabolic dysfunction in the female. *Mol Cell Endocrinol* 246: 165–174. https://doi. org/10.1016/j.mce.2005.11.016.
- Pansarasa O, D'Antona G, Gualea M, Marzani B, Pellegrino M, Marzatico F (2002) "Oxidative stress": effects of mild endurance training and testosterone treatment on rat gastrocnemius muscle. *Eur J Appl Physiol* 87: 550–555. https://doi.org/10.1007/s00421-002-0668-3.
- Pelletier J, Garnier DH, Reviers MM d, Terqui M, Ortavant R (1982) Seasonal variation in LH and testosterone release in rams of two breeds. *Reproduction* 64: 341–346. https://doi.org/10.1530/jrf.0.0640341.
- Pérez-Rodríguez L, Blas J, Viñuela J, Marchant TA, Bortolotti GR (2006) Condition and androgen levels: are condition-dependent and testosterone-mediated traits two sides of the same coin? *Anim Behav* 72: 97–103. https://doi.org/10.1016/j.anbehav.2005.09.021.
- Place NJ, Kenagy GJ (2000) Seasonal changes in plasma testosterone and glucocorticosteroids in free-living male yellow-pine chipmunks and the response to capture and handling. *J Comp Physiol B* 170: 245–251. https://doi.org/10.1007/s003600050282.
- Poole JH (1987) Rutting behavior in African elephants: the phenomenon of musth. *Behaviour* 102: 283–316. https://doi.org/10.1163/1568539 86X00171.
- Poole JH (1989) Announcing intent: the aggressive state of musth in African elephants. *Anim Behav* 37: 140–152. https://doi.org/10.1016/0003-3472(89)90014-6.
- Poole JH, Kasman LH, Ramsay EC, Lasley BL (1984) Musth and urinary testosterone concentrations in the African elephant (*Lox-odonta africana*). *Reproduction* 70: 255–260. https://doi.org/10.1530/ jrf.0.0700255.
- R Core Team (2022) R: A language and environment for statistical computing. *R Foundation for Statistical Computing*, Vienna, Austria. https://www.R-project.org/.
- Rasmussen LE, Buss IO, Hess DL, Schmidt MJ (1984) Testosterone and dihydrotestosterone concentrations in elephant serum and temporal

.....

gland secretions. *Biol Reprod* 30: 352–362. https://doi.org/10.1095/ biolreprod30.2.352.

- Rasmussen LEL, Hall-Martin AJ, Hess DL (1996) Chemical profiles of male African elephants, *Loxodonta africana*: physiological and ecological implications. *J Mammal* 77: 422–439. https://doi.org/10.2307/ 1382819.
- Rasmussen LEL, Perrin TE (1999) Physiological correlates of musth: lipid metabolites and chemical composition of exudates. *Physiol Behav* 67: 539–549. https://doi.org/10.1016/S0031-9384(99)00114-6.
- Reed WL, Clark ME, Parker PG, Raouf SA, Arguedas N, Monk DS, Snajdr E, Nolan V Jr, Ketterson ED (2006) Physiological effects on demography: a long-term experimental study of testosterone's effects on fitness. *Am Nat* 167: 667–683. https://doi.org/10.1086/ 503054.
- Robinson MR, Mar KU, Lummaa V (2012) Senescence and agespecific trade-offs between reproduction and survival in female Asian elephants. *Ecol Lett* 15: 260–266. https://doi.org/10.1111/ j.1461-0248.2011.01735.x.
- Ruiz M, French SS, Demas GE, Martins EP (2010) Food supplementation and testosterone interact to influence reproductive behavior and immune function in *Sceloporus graciosus*. *Horm Behav* 57: 134–139. https://doi.org/10.1016/j.yhbeh.2009.09.019.
- Rutkowska J, Cichoń M, Puerta M, Gil D (2005) Negative effects of elevated testosterone on female fecundity in zebra finches. *Horm Behav* 47: 585–591. https://doi.org/10.1016/j.yhbeh.2004.12.006.
- Saketos M, Sharma N, Santoro NF (1993) Suppression of the hypothalamic-pituitary-ovarian axis in normal women by glucocorticoids. *Biol Reprod* 49: 1270–1276. https://doi.org/10.1095/biolreprod49.6.1270.
- dos Santos DFJ, Berger V, Cristofari R, Htut W, Nyein UK, Aung HH, Reichert S, Lummaa V (2020) Seasonal variation of health in Asian elephants. *Conserv Physiol* 8: coaa119. https://doi.org/10.1093/ conphys/coaa119.
- Sapolsky RM (2002) Endocrinology of the stress-response. In JB Becker, SM Breedlove, D Crews, MM McCarthy, eds, *Becker JB, Breedlove SM, Crews D, McCarthy MM, eds. Behavioral Endocrinology*, Ed2nd. The MIT Press, Cambridge, MA, US, pp. 409–450.
- Schmidt MJ, Mar KU (1996) Reproductive performance of captive Asian elephants in Myanmar. *Gajah* 16: 23–42.
- Seltmann MW, Ukonaho S, Reichert S, Dos Santos D, Nyein UK, Htut W, Lummaa V (2020) Faecal glucocorticoid metabolites and H/L ratio are related markers of stress in semi-captive Asian timber elephants. *Animals* 10: 94. https://doi.org/10.3390/ani10010094.
- Seraphin S, Whitten PL, Reynolds VB (2008) The influence of age on fecal steroid hormone levels in male Budongo forest chimpanzees (*Pan troglodytes schweinfurthii*). Am J Primatol 70: 661–669. https:// doi.org/10.1002/ajp.20541.
- Shoshani J, Eisenberg JF (1982) Elephas maximus. *Mamm Species* 182: 1–8. https://doi.org/10.2307/3504045.

- Silverin B (1980) Effects of long-acting testosterone treatment on freeliving pied flycatchers, *Ficedula hypoleuca*, during the breeding period. *Anim Behav* 28: 906–912. https://doi.org/10.1016/S0003-3472(80))80152-7.
- Stoehr AM, Hill GE (2000) Testosterone and the allocation of reproductive effort in male house finches (*Carpodacus mexicanus*). *Behav Ecol Sociobiol* 48: 407–411. https://doi.org/10.1007/s002650000247.
- Sukumar R (2003) The Living Elephants: Evolutionary Ecology, Behavior, and Conservation. Oxford University Press, New York
- Sun B-J, Du W-G, Shu L, Chen Y, Wang Y (2011) The influence of thermal environment and food availability on testosterone and gonadal recrudescence in male Chinese skinks [*Plestiodon (Eumeces) chinensis*]. Gen Comp Endocrinol 170: 449–454. https://doi.org/10.1016/j. ygcen.2010.10.017.
- Supanta J, Brown JL, Bansiddhi P, Thitaram C, Punyapornwithaya V, Punturee K, Towiboon P, Somboon N, Khonmee J (2024) Physiological changes in captive elephants in northern Thailand as a result of the COVID-19 tourism ban—stress biomarkers. *Front Vet Sci* 11. https://doi.org/10.3389/fvets.2024.1351361.
- Tam NNC, Gao Y, Leung Y-K, Ho S-M (2003) Androgenic regulation of oxidative stress in the rat prostate: involvement of NAD(P)H oxidases and antioxidant defense machinery during prostatic involution and regrowth. Am J Pathol 163: 2513–2522. https://doi.org/10.1016/ S0002-9440(10)63606-1.
- Taya K, Komura H, Kondoh M, Ogawa Y, Nakada K, Watanabe G, Sasamoto S, Tanabe K, Saito K, Tajima H *et al.* (1991) Concentrations of progesterone, testosterone and estradiol-17β in the serum during the estrous cycle of Asian elephants (*Elephas maximus*). *Zoo Biol* 10: 299–307. https://doi.org/10.1002/zoo.1430100404.
- Taylor GT, Komitowski D, Muto S, Weiss J (1989) Behavioral endocrinology of male rats with periodic sexual contacts with exclusive or varied females. *Proc Soc Exp Biol Med* 192: 236–241. https://doi. org/10.3181/00379727-192-42991.
- Thongtip N, Saikhun J, Mahasawangkul S, Kornkaewrat K, Pongsopavijitr P, Songsasen N, Pinyopummin A (2008) Potential factors affecting semen quality in the Asian elephant (*Elephas maximus*). *Reprod Biol Endocrinol* 6: 9. https://doi.org/10.1186/1477-7827-6-9.
- Toro-Urrego N, Garcia-Segura LM, Echeverria V, Barreto GE (2016) Testosterone protects mitochondrial function and regulates Neuroglobin expression in astrocytic cells exposed to glucose deprivation. *Front Aging Neurosci* 8. https://doi.org/10.3389/fnagi.2016.00152.
- Túnez I, Feijóo M, Collado JA, Medina FJ, Peña J, Muñoz M d C, Jimena I, Franco F, Rueda I, Muntané J et al. (2007) Effect of testosterone on oxidative stress and cell damage induced by 3-nitropropionic acid in striatum of ovariectomized rats. *Life Sci* 80: 1221–1227. https://doi.org/10.1016/j.lfs.2006.12.013.
- Ukonaho S, Berger V, Franco dos Santos DJ, Htut W, Aung HH, Nyeing UK, Reichert S, Lummaa V (2023) Seasonal variation in molecular and physiological stress markers in Asian elephants. *Conserv Physiol* 11: coad029. https://doi.org/10.1093/conphys/coad029.

- Veiga JP, Polo V, Demas AEGE, Geber EMA (2008) Fitness consequences of increased testosterone levels in female spotless starlings. *Am Nat* 172: 42–53. https://doi.org/10.1086/587850.
- Vleck CM, Vertalino N, Vleck D, Bucher TL (2000) Stress, corticosterone, and heterophil to lymphocyte ratios in free-living Adélie penguins. *Condor* 102: 392–400. https://doi.org/10.1093/condor/102. 2.392.
- Watson R, Munro C, Edwards KL, Norton V, Brown JL, Walker SL (2013) Development of a versatile enzyme immunoassay for non-invasive assessment of glucocorticoid metabolites in a diversity of taxonomic species. *Gen Comp Endocrinol* 186: 16–24. https://doi.org/10.1016/j. ygcen.2013.02.001.
- Whitehead PE, McEwan EH (1973) Seasonal variation in the plasma testosterone concentration of reindeer and caribou. *Can J Zool* 51: 651–658. https://doi.org/10.1139/z73-096.
- Whitworth DJ, Licht P, Racey PA, Glickman SE (1999) Testis-like steroidogenesis in the ovotestis of the European mole, *Talpa europaea*. *Biol Reprod* 60: 413–418. https://doi.org/10.1095/biolreprod60. 2.413.
- Wickham H (2016) *Ggplot2: Elegant Graphics for Data Analysis.* Springer-Verlag, New York
- Williams C, (Group) ST (IUCN SAES), Varun Goswami (Conservation Initiatives I, Inc) S de S (Trunks & L), A Kumar (Nature Conservation Foundation I), N Baskaran (A. V.C. College TN, Pdr) KY (WWF L), Vivek Menon (Wildlife Trust of India and Chair ISAESG) (2019) IUCN red list of threatened species: *Elephas maximus*. *IUCN Red List Threat Species*.
- Wilson EK, Rogler JC, Erb RE (1979) Effect of sexual experience, location, malnutrition, and repeated sampling on concentrations of testosterone in blood plasma of *Gallus domesticus* roosters. *Poult Sci* 58: 178–186. https://doi.org/10.3382/ps.0580178.
- Wingfield JC (1984) Androgens and mating systems: testosteroneinduced polygyny in normally monogamous birds. *Auk* 101: 665–671. https://doi.org/10.2307/4086893.
- Wingfield JC (1988) Changes in reproductive function of free-living birds in direct response to environmental perturbations. In MH Stetson, ed, *Processing of Environmental Information in Vertebrates*. Springer, New York, NY, pp. 121–148
- Wingfield JC, Hegner RE, Dufty AM, Ball GF (1990) The "challenge hypothesis": theoretical implications for patterns of testosterone secretion, mating systems, and breeding strategies. *Am Nat* 136: 829–846. https://doi.org/10.1086/285134.
- Wingfield JC, Kenagy GJ (1991) Natural regulation of reproductive cycles. In PKT Pawg, MP Schreibruan, eds, *Vertebrate Endocrinology: Fundamentals and Biomedical Implications*. Academic Press, San Diego, pp. 181–241
- Wingfield JC, Lynn SE, Soma KK (2001) Avoiding the 'costs' of testosterone: ecological bases of hormone-behavior interactions. *Brain Behav Evol* 57: 239–251. https://doi.org/10.1159/000047243.

- Wingfield JC, Sapolsky RM (2003) Reproduction and resistance to stress: when and how. *J Neuroendocrinol* 15: 711–724. https://doi. org/10.1046/j.1365-2826.2003.01033.x.
- Yan W, Kang Y, Ji X, Li S, Li Y, Zhang G, Cui H, Shi G (2017) Testosterone upregulates the expression of mitochondrial ND1 and ND4 and alleviates the oxidative damage to the nigrostriatal dopaminergic system in orchiectomized rats. *Oxid Med Cell Longev* 2017: 1202459. https://doi.org/10.1155/2017/1202459.
- Yang Y, Tummaruk P, Angkawanish T, Langkaphin W, Chatdarong K (2023) Seasonal effects on body condition and characteristics of the estrous cycle in captive Asian elephants (*Elephas maximus*) in Thailand: a retrospective study. *Animals* 13: 1133. https://doi.org/10. 3390/ani13071133.
- Yon L, Chen J, Moran P, Lasley B (2008) An analysis of the androgens of musth in the Asian bull elephant (*Elephas maximus*). *Gen Comp Endocrinol* 155: 109–115. https://doi.org/10.1016/j.ygcen. 2007.03.006.

- Yon L, Kanchanapangka S, Chaiyabutr N, Meepan S, Stanczyk FZ, Dahl N, Lasley B (2007a) A longitudinal study of LH, gonadal and adrenal steroids in four intact Asian bull elephants (*Elephas maximus*) and one castrate African bull (*Loxodonta africana*) during musth and non-musth periods. *Gen Comp Endocrinol* 151: 241–245. https://doi. org/10.1016/j.ygcen.2007.01.028.
- Yon L, Kanchanapangka S, Chaiyabutr N, Stanczyk F, Meepan S, Lasley B (2007b) ACTH stimulation in four Asian bull elephants (*Elephas maximus*): an investigation of androgen sources in bull elephants. *Gen Comp Endocrinol* 151: 246–251. https://doi.org/10.1016/ j.ygcen.2007.01.029.
- Zhu X-D, Bonet B, Knopp RH (1997) 17β-estradiol, progesterone, and testosterone inversely modulate low-density lipoprotein oxidation and cytotoxicity in cultured placental trophoblast and macrophages. *Am J Obstet Gynecol* 177: 196–209. https://doi.org/10.1016/S0002-9378(97)70462-9.